

ANALYSIS OF MARINE PHYTOPLANKTON IN THE YODO RIVER ESTUARY BY THE NUMERICAL ECOSYSTEM MODEL

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In order to understand the temporal variation of the physics and fluid structure of Yodo River estuary in detail, we had made in-situ observation. And the temporal variation of *Alexandrium tamarense* which cause the shellfish poisoning of natural freshwater clam was analyzed by the numerical ecosystem model which is considered the salinity effects. Stratification develops in the downstream side. Chl.a concentration is high in the seawater region. *A. tamarense* is detected in the downstream side. The numerical ecosystem model including the salinity effect for *A. tamarense* was formulated. *A. tamarense* grow only in the bottom layer in daytime, and the daily mean of it is 7 % of it transported from Osaka Bay. *A. tamarense* is transported to the upstream in flood tide. 81 % of it transported from Osaka Bay goes to the upstream zone. Much *A. tamarense* transported to the upstream zone in nighttime due to the vertical migration. Therefore when it is the flood tide in nighttime, more of *A. tamarense* might be transported to the upstream zone.

Key words: Alexandrium tamarense, Yodo River, estuary, numerical ecosystem model, red tide, shellfish poisoning

I. INTRODUCTION

The Yodo River estuary shown in Fig. 1 which is defined in this research as a zone from the river mouth up to 10 km, has fisheries of natural freshwater clam around at 5 km from the river mouth. Shellfish poisoning of natural freshwater clam occurred intermittently in the estuary in spring since 2007, and the shipment of freshwater clams was halted due to government regulations. Cause of the shellfish poison is *Alexandrium tamarense* [1] which is poisonous marine phytoplankton appearing in Osaka Bay every year, and can growth in salinity environment of more than 15 [2]. The lack of fresh water discharge from the weir promoted the initiation of the bloom of *A. tamarense* [3]. In order to understand the temporal variation of the physics and fluid structure of the area in detail, we made in-situ observation and analyzed the temporal variation of *A. tamarense* by using the numerical ecosystem model.

II. METHODS

The observation was carried out during April 2nd and 3rd in 2012 in the neap tide. Vertical profiles of water temperature (T), salinity (S), fluorescence, photon (I) and so on were measured by CTD (the conductivity, temperature and depth profiler) in 10 cm width at 5 stations in the intertidal zone between the river mouth and the Yodo River weir shown in Fig. 1. Surface water (0 m) and

the bottom water which is above 1 m from the bottom were sampled at the same time by a bucket and Kitahara water sampler, respectively. Sampled water was dispensed to the poly bottles, and was analyzed for the chemical and biological analysis. The items are cell density of *A. tamarensis*, Chl.a (chlorophyll-a), TP (total phosphorus) and DIP (dissolved inorganic phosphorous) concentrations and so on. Sampled water were kept in the dark and cool condition, and were transported to Research Institute of Environment, Agriculture and Fisheries, Osaka Prefecture to count the cell density, and to Hyogo Environmental Advancement Association to analyze Chl.a, TP and DIP concentrations and so on. The fluorescence values of CTD were converted into Chl.a concentrations using the analyzed Chl.a concentrations. ADCP (Acoustic Doppler Current Profiler) observation was carried out along the traverse line (L1 in Fig. 1) at the river mouth in every 23 cm depth at 0.2 Hz. The observation was repeated 4 cycles during one tidal period shown in Fig. 2(a). The water level was monitored every 10 minutes at Fukushima as shown in Fig. 1 by the Yodogawa River Office, Kinki Regional Development Bureau, Ministry of Land, Infrastructure and Transport.

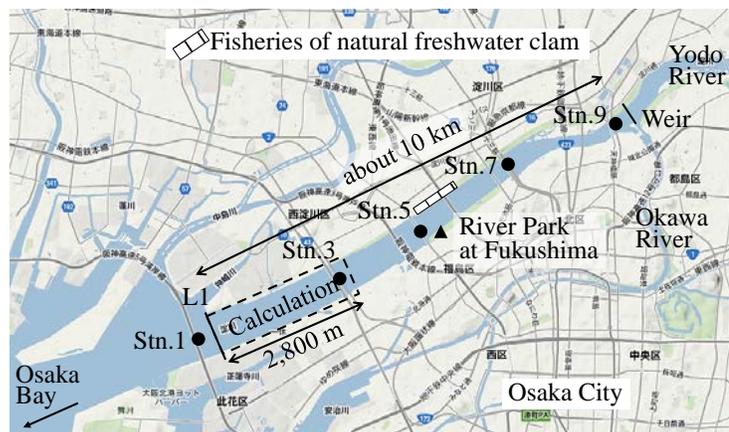
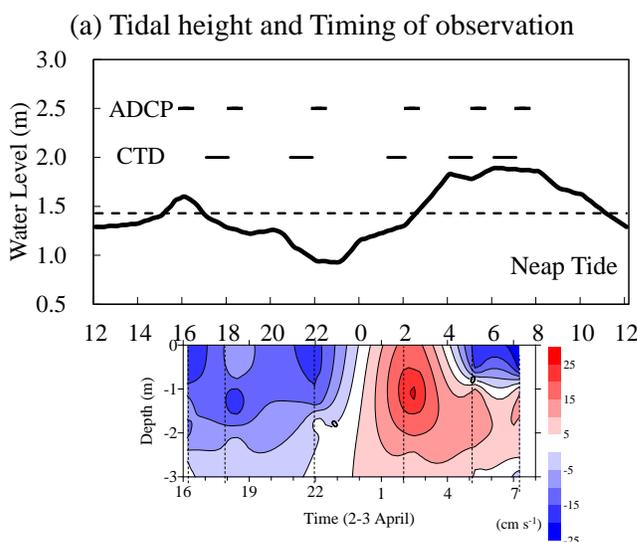


Fig. 1. Study area.



(b) Flow velocity at the river mouth.

Fig. 2. Temporal variations of tidal change.

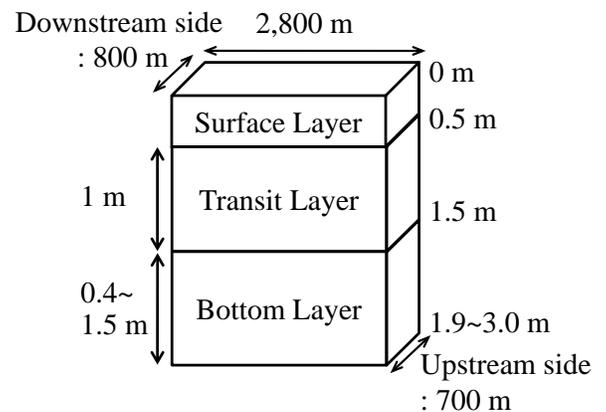


Fig. 3. Size of the calculation zone.

The calculation zone by the numerical ecosystem model shown in Fig. 1 is from L1 up to 2,800 m upstream. Water depth became deeper toward the upstream side, and is shallow in the outside of the upstream boundary. Stn. 3 is included in the zone. The calculation zone was divided into the surface layer (above 0.5 m), the transition layer (0.5-1.5 m) and the bottom layer (below 1.5 m) as shown in Fig. 3. The phosphorus (P) cycling during 24 hours, which is one tidal period of the day was calculated. DIN/DIP ratio is larger than the Redfield ratio (C:N:P=106:16:1) in this observation. It means that the primary production is limited by DIP. The compartments of the numerical ecosystem model is shown in Fig. 4. The types of P are DIP, DOP, phytoplankton, *A. tamarensis*, zooplankton and detritus, and are changed by the biochemical processes. Phytoplankton in this research means other than *A. tamarensis*, and is mainly diatom. *A. tamarensis* utilizes DOP in photosynthesis. The exchange processes between the layers or the outside of the area are the horizontal and vertical advection, the vertical diffusion, the natural sinking of phytoplankton and detritus and the diurnal vertical migration of *A. tamarensis*. *A. tamarensis* move to the surface during the day, and move to the bottom during the night. The horizontal diffusion would be quite small compare to the horizontal advection. Therefore the horizontal diffusion is not assumed. The water depth is shallow in the river. So, the vertical advection speed is assumed as a constant in each depth. The upstream and upward fluxes are positive.

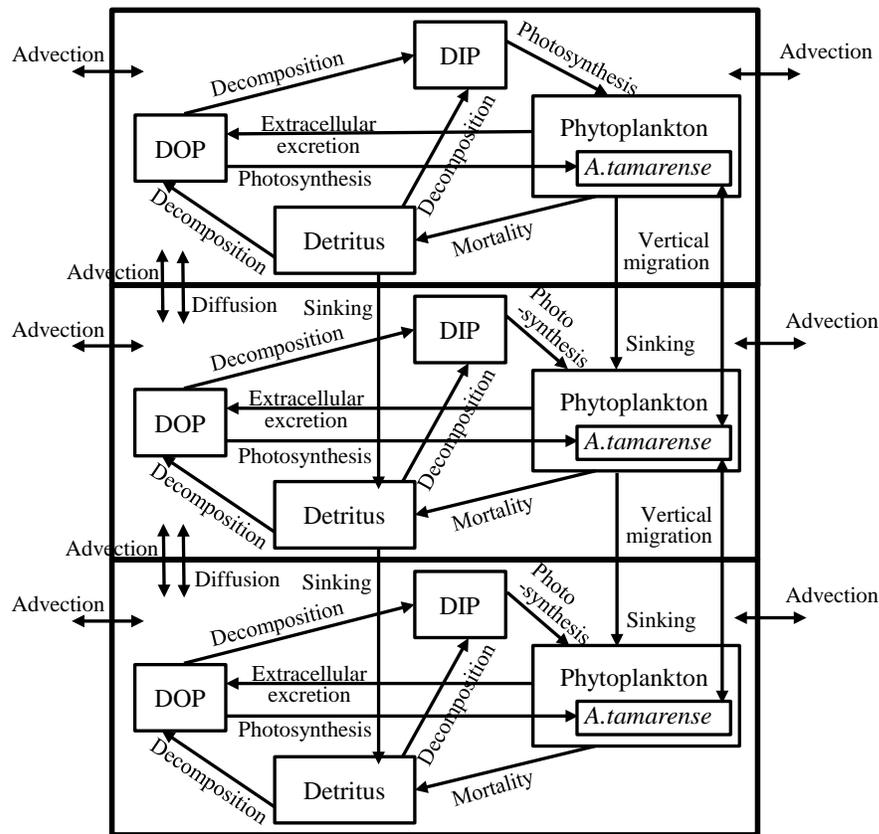


Fig. 4. The compartments of the numerical ecosystem model

Temporal changes in P concentration of each compartment, P in phytoplankton, *A. tamarensis*, zooplankton and detritus, DIP and DOP, are represented by the equations. For example,

equation (1) represents the temporal change of P concentration in *A. tamarensis* (*ATM*). The first and second terms mean photosynthesis. The 3rd term means extra cellular excretion of DOP by photosynthesis. The 4th term means mortality. The 5th term means grazing by zooplankton. The 6th term means the horizontal and vertical transports including the vertical migration.

$$\begin{aligned} \frac{dATM}{dt} = & A_{1-i}ATM_0 + A_{1-o}ATM_0 - A_2(A_{1-i} + A_{1-o})ATM_0 - A_3ATM_0 - B_1ZOO_0 \\ & + \frac{1}{V_0}(F_dU_dATM_d - F_uU_uATM_u + F_sWATM_v + F_sw_tATM_0 + F_s\frac{K}{H_v}(ATM_v - ATM_0)) \end{aligned} \quad (1)$$

where A 's are the coefficients of biochemical processes related to phytoplankton, and will be explained later. B_1 represents grazing by zooplankton. Subscripts of the concentrations mean place. Subscript 0 refers to the calculating layer. Subscript d and u refer to the downstream and upstream side, respectively. But when the water flows out from the calculation layer, the concentration of the calculated layer should be used. Subscript v refers to the source layer of outflow. V_0 is the volume of the calculation layer. H_v is the distance between the surface and transition or the transition and bottom layers. F 's are the section. U 's are the horizontal advection velocity. Subscript d refers to the downstream side, u is the upstream side, and s is the surface. W is the vertical advection speed. K is the vertical eddy diffusivity. The estimation method of U , W and K will be explained later. w_t is the vertical migration speed of *A. tamarensis*. It is $w_t > 0$ during the day (at 06-18), and means the upward motion. It is $w_t < 0$ during the night (at 18-06), and means the downward motion. Moreover, *A. tamarensis* would not swim to the lower salinity layer. Therefore, the vertical migration is stopped not to move to the layer that salinity is below the threshold. The terms of vertical advection, diffusion and migration are double in the transition layer which have two boundaries with the upper and lower sides.

A_1 represents the photosynthesis, and is the function of DIP or DOP concentration (DIP_0 , DOP_0), T , I and S . Equation (2) shows in case of DIP. A_1 of phytoplankton does not include the term of salinity. And phytoplankton does not use DOP in photosynthesis.

$$A_{1-i} = V_{max} \cdot \frac{DIP_0}{DIP_0 + k_p} \cdot \frac{T}{T_o} \exp(1 - \frac{T}{T_o}) \cdot \frac{I}{I_o} \exp(1 - \frac{I}{I_o}) \cdot [1 - \exp\{-k_{gs}(S^* - S)\}] \quad (2)$$

The term of nutrient represents by the Michaelis-Menten equation. V_{max} is the maximum uptake ratio of nutrient. k_p is the half saturation constant of nutrient for phytoplankton. V is V_{max} and k_p are different depending on the type of nutrients and the phytoplankton species. Shapes of the temperature and photon terms represent that growth is most active in the optimum water temperature (T_o) and photon (I_o). However photoinhibition do not occur even when the photon in the water is more than I_o . So, $I=I_o$ is applied when $I > I_o$. The function of salinity effect is formulated based on the result of incubation shown in Fig. 5 [2]. And the threshold of salinity (S_g^*) which stop the growth of *A. tamarensis*, and the coefficient of salinity dependency in the photosynthesis of *A. tamarensis* (k_{gs}) are given.

A_2 is the coefficient of the extra cellular excretion of DOP by photosynthesis. A_3 represents the mortality, and is given by the following equation which is the function of T , and of S only case of *A. tamarensis*. The function of salinity effect is also formulated by Fig. 5.

$$A_3 = m_{p0} \exp(k_{m-p}T) \exp\{k_{ms}(S_m^* - S)\} \quad (3)$$

where m_{p0} is the mortality speed of phytoplankton at 0 deg-C, k_{m-p} is the temperature dependency of mortality for phytoplankton, k_{ms} is the salinity dependency of mortality for *A. tamarensis*, S_m^* is the threshold of salinity in photosynthesis of *A. tamarensis*. Mortality accelerates when salinity is over S_m^* .

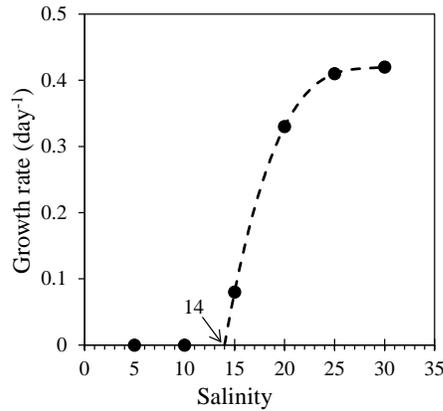


Fig. 5. Growth rate of *A. tamarensis* [2].

Parameters of constant of the ecosystem model are shown in Table 1, and are referred to the previous study [4]. It analyzed the competitive relationship of diatom and non-diatom in the inner part of Osaka Bay which is the estuary of the Yodo River. w_t has been tuned to reproduce the observed value, and is 1/20 of the value by reference [4]. V_{max} and k_p related to photosynthesis are referred to previous study [5] based on the incubation by using the Yodo River water. The parameters of salinity for *A. tamarensis* were determined based on Fig. 5.

The data sets of the boundary and environmental conditions of T , S , I , V , H_v and F_v , which vary temporally and are inputted to the model every minute for 24 hours, were constructed by the observation data. The variations of salinity are 1.0-2.2 in the surface layer, 9.1-12.7 in the transition layer and 24.6-26.5 in the bottom layer. Salinity in the surface and transition layer are always less than 14 which is the threshold of salinity in photosynthesis of *A. tamarensis*. The variations of water temperature is 10.8-11.6 deg-C which is lower than the optimum water temperature. Photoenvironments were the optimum condition at 6 to 18 in the surface layer and at 8 to 16 in the transition layer. Photon in the bottom layer was 7 % of it in the surface layer, and is enough for photosynthesis.

P concentrations in Stn. 1 were used as the boundary condition of the downstream side, and it in Stn. 5 was used as the upstream side. P concentrations in phytoplankton were calculated by Chl.a concentration converted from fluorescence values of CTD and P/Chl.a which was estimated by C/Chl.a=30 [6], the Redfield ratio and the atomic weight of C and P. P concentrations in

zooplankton were assumed 10 % of phytoplankton. P concentrations in *A. tamarensis* estimated by the cell density and P weigh par cell which calculated by C weigh per cell and the Redfield ratio. 2,000 pg-C cell⁻¹ [7] was used, then 48.7 pg-P cell⁻¹ was obtained. TP:DOP in the inter tidal zone of Yodo Rivre is 14.5 % [8]. P concentrations in detritus is obtained by subtracting these concentrations from the TP.

Table 1. Values of the parameter used in the ecosystem model.

Parameters	Symbol	Value	unit
Surface Area of Box	F_s	2,100,000	m ³
Cross Section Area of the Upper Layer at the Downstream Boundary	F_d	400	m ²
Cross Section Area of the Middle Layer at the Downstream Boundary	F_d	800	m ²
Cross Section Area of the Upper Layer at the Upstream Boundary	F_u	350	m ²
Cross Section Area of the Middle Layer at the Upstream Boundary	F_u	700	m ²
Sinking Speed of Phytoplankton	w_p	23.2×10^{-5}	cm s ⁻¹
Sinking Speed of Detritus	w_d	23.2×10^{-4}	cm s ⁻¹
Vertical Migration Speed of <i>A. tamarensis</i>	w_t	0.39×10^{-3}	cm s ⁻¹
Maximum Uptake rate of Phytoplankton	V_{max-p}	0.75	day ⁻¹
Maximum Uptake rate of <i>A. tamarensis</i>	V_{max-t}	0.5	day ⁻¹
Half Saturation Constant of DIP for Phytoplankton	k_{p-p}	0.12	μM
Half Saturation Constant of DIP for <i>A. tamarensis</i>	k_{p-i}	0.15	μM
Half Saturation Constant of DOP for <i>A. tamarensis</i>	k_{p-o}	0.1	μM
Optimum Ligth Intensity of Photosynthesis for Phytoplankton	I_{o-p}	75	E m ² s ⁻¹
Optimum Ligth Intensity of Photosynthesis for <i>A. tamarensis</i>	I_{o-t}	100	E m ² s ⁻¹
Optimum water temerature of photosynthesis	T_o	30.8	°C
Salinity Dependency of Photosynthesis for <i>A. tamarensis</i>	k_{gs}	0.23	
Threshold of Salinity for Photosynthesis of <i>A. tamarensis</i>	S_g^*	14	psu
Ratio of extra cellular excretion of DOP by photosynthesis	A_2	0.135	g m ⁻¹ s ⁻¹
Mortality Speed of Phytoplankton at 0 deg-C	m_{p0}	8.0	m ³ g-P day ⁻¹
Temperature Dependency of Mortality for Phytoplankton	k_{m-p}	0.069	°C
Salinity Dependency of Mortality for <i>A. tamarensis</i>	k_{sm}	1.2	
Threshold of Salinity for Mortality of <i>A. tamarensis</i>	S_m^*	14	psu
Ivlev Constant	λ	0.47	
Threshold of Phytoplankton Density for Grazing by Zooplankton	PHP^*	0.1	mg Chl.a ⁻¹
Grazing Speed of Phytoplankton by Zooplankton at 0 deg-C	g_{p0}	0.1	
Temperature Dependency of Grazing for Zooplankton	k_{g-p}	0.069	°C ⁻¹
Constant for Urine Generation of Zooplankton	α	0.4	
Constant for Fecal Pellet Generation of Zooplankton	β	0.3	
Mortality Speed of Zooplankton at 0 deg-C	m_{z0}	30.0	
Temperature Dependency of Mortality for Zooplankton	k_{m-z}	0.069	°C ⁻¹
Decomposition Speed of Detritus to DIP at 0 deg-C	d_{di}	0.03	
Decomposition Speed of Detritus to DOP at 0 deg-C	d_{do}	0.03	
Decomposition Speed of DOP to DIP at 0 deg-C	d_{oi}	0.03	
Temperature Dependency of Decomposition of Detritus to DIP	k_{di}	0.069	°C ⁻¹
Temperature Dependency of Decomposition of Detritus to DOP	k_{do}	0.069	°C ⁻¹
Temperature Dependency of Decomposition of DOP to DIP	k_{oi}	0.069	°C ⁻¹

U , W and K_v are estimated by the physical model. The governing equations based on water and salt balances. The water balance in each layer are given by the following equations.

$$\begin{aligned}\frac{dV_1}{dt} &= U_{d1}F_{d1} - U_{u1}F_{u1} + WA_s = 0 \\ \frac{dV_2}{dt} &= U_{d2}F_{d2} - U_{u2}F_{u2} = 0 \\ \frac{dV_3}{dt} &= U_{d3}F_{d3} - U_{u3}F_{u3} - WF_s = \frac{dH_3}{dt}F_s\end{aligned}\quad (4)$$

where subscript u refers to the upstream boundary, d is the downstream boundary. The subscripts of number refer to the layer. H_3 is the thickness of the bottom layer. Since V_1 and V_2 are constant, the water budgets by advection are zero. The variation of V_3 by the tidal change and the budget of advection are balanced in the bottom layer. U_{d1} , U_{d2} and U_{d3} were obtained by ADCP observation. Therefore, the unknown quantities are U_{u1} , U_{u2} , U_{u3} and W .

The salt balance in each layer are given by the following equations.

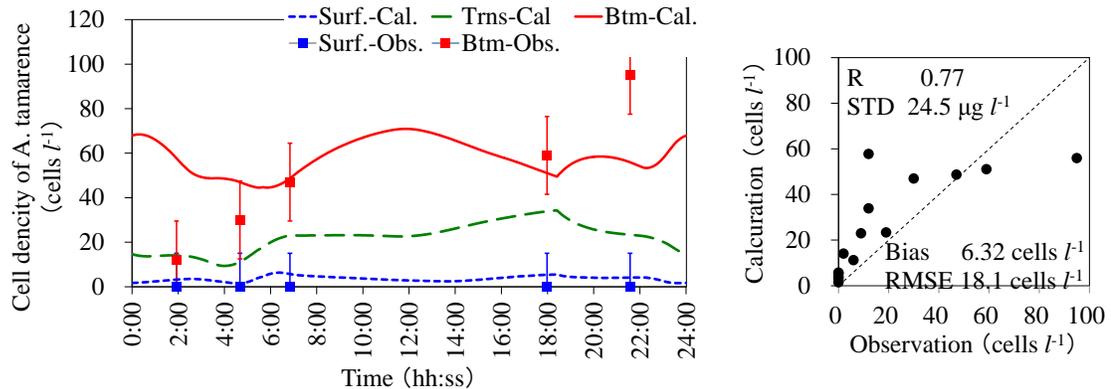
$$\begin{aligned}\frac{dS_1}{dt}H_1F_s &= U_{d1}S_{d1}F_{d1} - U_{u1}S_{u1}F_{u1} + WS_2F_s + K_{v12}\frac{S_2 - S_1}{H_{12}}F_s \\ \frac{dS_2}{dt}H_2F_s &= U_{d2}S_{d2}F_{d2} - U_{u2}S_{u2}F_{u2} - WS_2F_s + WS_3F_s + K_{v12}\frac{S_{01} - S_{02}}{H_{12}}F_s + K_{v23}\frac{S_{03} - S_{02}}{H_{23}}F_s \\ \frac{dS_3}{dt}H_3F_s &= U_{d3}S_{d3}F_{d3} - U_{u3}S_{u3}F_{u3} - WS_3F_s + K_{v23}\frac{S_{02} - S_{03}}{H_{23}}F_s\end{aligned}\quad (5)$$

where subscript 12 refers to the boundary of the surface and transition layers, 23 is the boundary of the transition and bottom layers. The additional unknown quantities are K_{v12} and K_{v23} . The unknown quantities at the observation time are obtained by the inverse matrix because there are six equations for six unknown quantities.

III. RESULTS AND DISCUSSIONS

Stratification develops in the downstream side. Seawater intrudes to close to Stn.9 in flood and high tides. And Chl.a concentration is high in the seawater region. Seawater intrusion stops at around Stn.7 in ebb and low tides. High concentration of Chl.a near the surface would be freshwater phytoplankton. Seawater expansion to the river spread a habitat of marine phytoplankton [9]. *A. tamarensis* is detected in the downstream side mainly, therefore the calculation zone has been set at the river mouth. However, the cell density is small, and the red tide was not formulated in this year. Figure 2(b) shows the depth-time cross section of the flow velocity at the river mouth. Plus means the upstream flow. It is the downstream flow in all depth from the observation start until 23 o'clock which is weak ebb tide. It is the upstream flow in all depth from 1 to 4 o'clock which is flood tide. Since the variations of horizontal advection by the budget estimates, it is the downstream flow in all boundaries during 13-21. Therefore, this period was defined as ebb tide. And the other period,

during 21-13, was defined as flood tide. The estimated vertical advection velocity is $\pm 9 \times 10^{-3} \text{ cm s}^{-1}$, and the vertical diffusion coefficient is $10^{-5} - 10^{-4} \text{ cm s}^{-2}$.



(a) Temporal variations of calculation and observation (b) Correlation with statistical values.

Fig. 6. Calculation result of *A. tamarensis*.

Figure 6(a) is the results of cell density of *A. tamarensis*. Dots show the observed data with the error bar which means the conversion error from concentration in cell density. The results almost reproduced the observed values. Figure 6(b) shows the correlation chart with statistical values. DIP and Chl.a concentrations are under estimate because freshwater phytoplankton is not considered in the model. But *A. tamarensis* is over estimate a little especially in the surface layer. Phytoplankton tends to increase only in the daytime in the bottom layer. Since *A. tamarensis* move to the bottom during night, the increase tendency is also in the nighttime a little.

Figure 7 shows the budgets of *A. tamarensis*. Each number is total flux of 24 hours, and has been standardized by the horizontal advection, $4,185 \times 10^3 \text{ mg}$, to the bottom layer of the downstream boundary. The standard number means the volume of *A. tamarensis* supplied from Osaka Bay. *A. tamarensis* grow only in the bottom layer, and the volume is 7 % of it from Osaka Bay (the numbers at the left side with an underline means photosynthesis in Fig. 8). *A. tamarensis* dies at the surface layer (the numbers at the right side with an underline means mortality & grazing), and moves to the bottom layer by the diurnal migration (the numbers of bold type means diurnal migration). These are salinity effect. 78 % of *A. tamarensis* transported from Osaka Bay to the bottom layer goes to the upstream zone (the numbers without decorations means advection). And 26% returns to the transit layer. 52% of *A. tamarensis* transported from Osaka Bay is lost in the upstream zone. *A. tamarensis* is increased in the bottom layer (the numbers in parentheses mean the budget). It suggests the long term variation, for example, spring and neap tide, and the seasonal variation.

Figure 8 shows the budgets which is divided to flood and ebb tides. Each number has been also standardized, and the volume in flood tide, $4,185 \times 10^3 \text{ mg}$, was used in this case. *A. tamarensis* is transported from Osaka Bay to the upstream zone in flood tide mainly (81%), and the half of this is returned in ebb tide (27+13 %). Other half is lost in the upstream zone. Return volume to the transit layer is larger than to the bottom layer. *A. tamarensis* would not go to lower salinity layer by itself. So, it is considered that *A. tamarensis* will be transported by the vertical advection and/or diffusion. Figure 9 shows the budgets which is divided to the daytime and nighttime. Each number has been also standardized, and the volume in daytime, $1,878 \times 10^3 \text{ mg}$, was used in this case. *A.*

tamarensis grow only in daytime in the bottom layer, and the volume is 16 % of it from Osaka Bay. *A. tamarensis* do not go to the transit and the surface layers by itself even daytime because of low salinity, but is transported by the vertical advection (12+22 %) and diffusion (15+32 %). Therefore *A. tamarensis* died at the surface layer (17 % & 22 %), and transported not so much to the upstream zone (27 %). *A. tamarensis* is also transported by the vertical advection (17+29 %) and diffusion (34+109 %) in nighttime. However the comparable volume (24 % & 139 %) move to the bottom layer by itself. Therefore much *A. tamarensis* (148 %) transported from the ocean go through upstream.

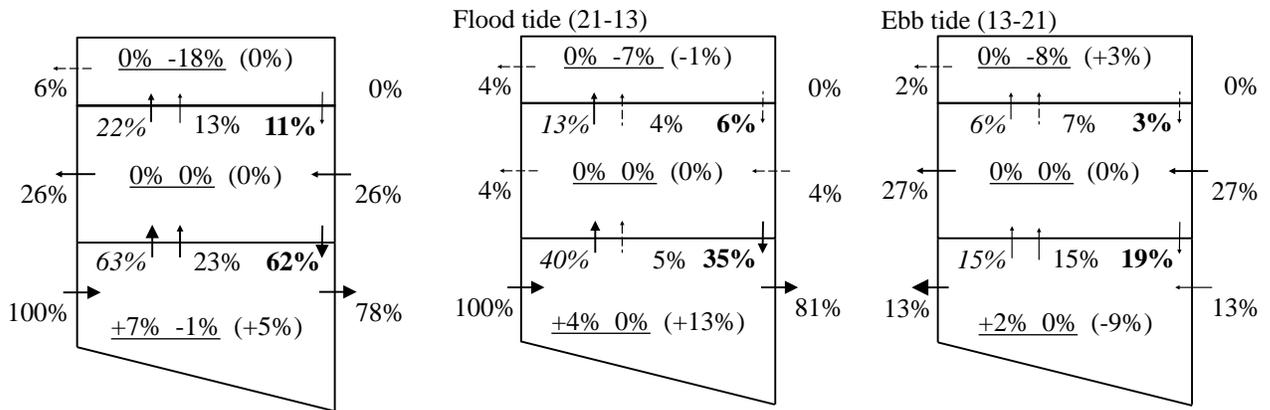


Fig. 7. Budgets of *A. tamarensis*. Fig. 8. Budgets of *A. tamarensis* divided to flood and ebb tides.

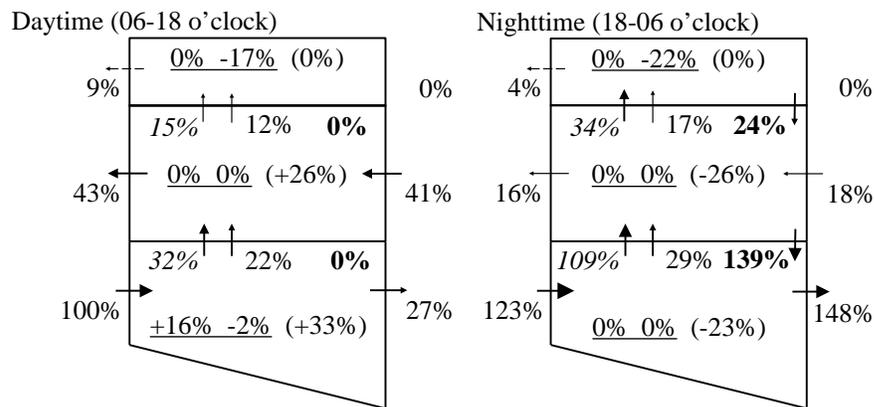


Fig. 9. Budgets of *A. tamarensis* divided to daytime and nighttime.

IV. CONCLUSION

In order to understand the temporal variation of the physics and fluid structure of Yodo River estuary in detail, we had made in-situ observation. And the temporal variation of *A. tamarensis* which cause the shellfish poisoning of natural freshwater clam was analyzed by the numerical ecosystem model which is considered the salinity effects.

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